

# List of Publications

## A. Publication During the Calendar year 2022

### • Department of Biological Sciences

1. **Aishwarya Agarwal and Samrat Mukhopadhyay (2022).** Prion Protein Biology Through the Lens of Liquid-Liquid Phase Separation: Liquid-liquid phase separation of prion protein. *Journal of Molecular Biology*, 434(1), 167368. <https://doi.org/10.1016/j.jmb.2021.167368>
2. **Aishwarya Agarwal, Lisha Arora, Sandeep K. Rai, Anamika Avni, and Samrat Mukhopadhyay (2022).** Spatiotemporal modulations in heterotypic condensates of prion and  $\alpha$ -synuclein control phase transitions and amyloid conversion. *Nature Communications*, 13(1), 1154. <https://doi.org/10.1038/s41467-022-28797-5>
3. **Aiswarya Sajeevan, Rakesh Pandian, and Shravan Kumar Mishra (2022).** Vectors with a flexible multiple cloning site and modular epitope tags for gene expression studies in *Schizosaccharomyces pombe*. *Gene Reports*, 29(1), 101681. <https://doi.org/10.1016/j.genrep.2022.101681>
4. **Alok Tiwary, Rahul Babu, Ruchira Sen, and Rhitoban Ray choudhury (2022).** Bacterial supergroup-specific “cost” of Wolbachia infections in *Nasoniavitripennis*. *Ecology and Evolution*, 12(9), 9219. <https://doi.org/10.1002/ece3.9219>
5. **Amjadudheen Varikka Pulakkal, Anuraag Ghosh, and Shravan Kumar Mishra (2022).** Broader roles of the ubiquitin-like protein Hub1 indicated by its yeast two-hybrid interactors. *MicroPublication Biology*, 35098049, 2022. <https://doi.org/10.17912/micropub.biology.000519>

6. **Anamika Avni, Aishwarya Agarwal, Sandeep K.Rai, Ashish Joshi, Anuja Walimbe, Swastik G. Pattanashettyand Samrat Mukhopadhyay (2022).** Single-droplet vibrational raman spectroscopy illuminates the inner workings of phase-separated biomolecular condensates. *Biophysical Journal*, 121(3), 307-308. <https://doi.org/10.1016/j.bpj.2021.11.1216>
7. **Anamika Avni, Ashish Joshi, Anuja Walimbe, Swastik G. Pattanashetty, and Samrat Mukhopadhyay (2022).** Single-droplet surface-enhanced Raman scattering decodes the molecular determinants of liquid-liquid phase separation. *Nature Communications*, 13(1), 4378. <https://doi.org/10.1038/s41467-022-32143-0>
8. **Anamika Kumari, Anjali Yadav, and Indrajit Lahiri (2022).** Transient State Kinetics of Plasmodium falciparum Apicoplast DNA Polymerase Suggests the Involvement of Accessory Factors for Efficient and Accurate DNA Synthesis. *Biochemistry*, 61(21), 2319-2333. <https://doi.org/10.1021/acs.biochem.2c00446>
9. **Anish Kumar Mondal and Kausik Chattopadhyay (2022).** Structures and functions of the membrane-damaging pore-forming proteins. *Advances in Protein Chemistry and Structural Biology*, 128(1), 241-288. <https://doi.org/10.1016/bs.apcsb.2021.07.001>
10. **Anish Kumar Mondal, Kusum Lata, Mahendra Singh, Shamaita Chatterjee, Aakanksha Chauhan, Sindhoora Puravankara and Kausik Chattopadhyay (2022).** Cryo-EM elucidates mechanism of action of bacterial pore-forming toxins. *Biochimica et Biophysica Acta - Biomembranes*, 1864(11), 184013. <https://doi.org/10.1016/j.bbamem.2022.184013>
11. **Anish Kumar Mondal, Nayanika Sengupta, Mahendra Singh, Rupam Biswas, Kusum Lata, Indrajit Lahiri, Somnath Dutta and Kausik Chattopadhyay (2022).** Glu289 residue in the pore-forming motif of Vibrio cholerae cytolsin is important for efficient β-barrel pore formation. *Journal of Biological Chemistry*, 298(10), 102441. <https://doi.org/10.1016/j.jbc.2022.102441>
12. **Anupa Majumdar and Samrat Mukhopadhyay (2022).** Excitation energy migration to study protein oligomerization and amyloid formation. *Biophysical Chemistry*, 281(1), 106719. <https://doi.org/10.1016/j.bpc.2021.106719>

13. **Anupa T Anil, Karan Choudhary, Rakesh Pandian, Praver Gupta, Poonam Thakran, Arashdeep Singh, Monika Sharma, and Shravan Kumar Mishra (2022).** Splicing of branchpoint-distant exons is promoted by Cactin Tls1 and the ubiquitin-fold-activated Sde2. *Nucleic Acids Research*, 50(17), 10000-10014. <https://doi.org/10.1093/nar/gkac769>
14. Apoorva Pandey, **Riya Madan**, and Swati Singh (2022). Immunology to Immunotherapeutics of SARS-CoV-2: Identification of Immunogenic Epitopes for Vaccine Development. *Current Microbiology*, 79(10), 30033. <https://doi.org/10.1007/s00284-022-03003-3>
15. **Archit Gupta, Ashish Joshi, Samrat Mukhopadhyay, and PurnanandaGuptasarma (2022).** The Escherichia coli nucleoid exists in a liquid-liquid phase separated state which seems to be responsive and tunable towards stresses. *Biophysical Journal*, 121(3), 357a. <https://doi.org/10.1016/j.bpj.2021.11.971>
16. **Arpita Mrigwani, Bhishem Thakur, Harman Kaur andPurnanandaGuptasarma (2022).** Synergistic action of different thermostable esterases in pet degradation: improvement of activity against intermediate degradation products by protein engineering. *Biophysical Journal*, 121(3), 347a-348a. <https://doi.org/10.1016/j.bpj.2021.11.1032>
17. **Arpita Mrigwani, BhishemThakura, and PurnanandaGuptasarma (2022).** Conversion of polyethylene terephthalate into pure terephthalic acid through synergy between a solid-degrading cutinase and a reaction intermediate-hydrolysing carboxylesterase. *Green Chemistry*, 24(17), 6707-6719. <https://doi.org/10.1039/d2gc01965e>
18. **Arpita Sarkar, Pallavi kaila Sharma, and PurnanandaGuptasarma(2022).** Exploring the novel mechanistic aspects of function of a hyperthermophile two-site exo-amylase-cum-glucanotransferase displaying substrate versatility. *Biophysical Journal*, 121(3), 346a. <https://doi.org/10.1016/j.bpj.2021.11.1025>
19. **Arpita Sharma, Shashi Prakash Yadav, Dwipjyoti Sarma and Arunika Mukhopadhyaya (2022).** Modulation of host cellular responses by gram-negative

- bacterial porins. *Advances in Protein Chemistry and Structural Biology*, 128(1), 35-77. <https://doi.org/10.1016/bs.apcsb.2021.09.004>
20. **Deeksha Thakur and Shashi B. Pandit (2022).** Substrate promiscuity: a continuum feature of enzymes. *Biophysical Journal*, 121(3), 344a. <https://doi.org/10.1016/j.bpj.2021.11.1015>
21. **Deeksha Thakur and Shashi Bhushan Pandit (2022).** Unusual commonality in active site structural features of substrate promiscuous and specialist enzymes. *Journal of Structural Biology*, 214(1), 107835. <https://doi.org/10.1016/j.jsb.2022.107835>
22. **Deepinder Kaur, Pratima Verma, Mahendra Singh, Arpita Sharma, Kusum Lata, Arunika Mukhopadhyaya, and Kausik Chattopadhyay (2022).** Pore formation-independent cell death induced by a  $\beta$ -barrel pore-forming toxin. *FASEB Journal*, 36(10), 2200788R. <https://doi.org/10.1096/fj.202200788R>
23. Deepjyoti Kumar Das, Mohammad Adeel Zafar, Sidhanta Nanda, Sanpreet Singh, Taruna Lamba, Hilal Bashir, Pargat Singh, Sudeep Kumar Maurya, Sajid Nadeem, SharvanSehrawat, Vijayender Bhalla, and Javed Naim Agrewala(2022). Targeting dendritic cells with TLR-2 ligand-coated nanoparticles loaded with Mycobacterium tuberculosis epitope induce antituberculosis immunity. *Journal of Biological Chemistry*, 298(12), 102596. <https://doi.org/10.1016/j.jbc.2022.102596>
24. **Devang Haresh Liya, NithishwerMouroug Anand, Ashwin Kumar Jainarayanan, MirudulaElanchezhian, Madhumati Seetharaman, DhanuushBalakannan, and Arpit Kumar Pradhan (2022).** Drug repurposing and sequence analysis in S-glycoprotein variants reveals critical signature patterns and destabilization of receptor-binding domain in omicron variant. *Journal of Biomolecular Structure and Dynamics*, 2127902. <https://doi.org/10.1080/07391102.2022.2127902>
25. **Garima Prazapati Ankit Yadav Anoop Ambili Abhilasha Sharma and RhitobanRaychoudhury (2022).** Males of the parasitoid wasp *Nasonia vitripennis* can identify which fly hosts contain females. *Royal Society Open Science*, 9(1), 211865. <https://doi.org/10.1098/rsos.211865>

26. **Garima Prazapati, Ankit Yadav, Anoop Ambili, Abhilasha Sharma and Rhitoban Raychoudhury (2022).** Males of the parasitoid wasp, *Nasonia vitripennis*, can identify which fly hosts contain females. *Royal Society Open Science*, 9(1). <https://doi.org/10.1098/rsos.211865>
27. Gaurav Kumar, **Prateek Chawla, Neha Dhiman**, Sanya Chadha, Sheetal Sharma, Kanupriya Sethi, **Mahak Sharma** and Amit Tuli (2022). RUFY3 links Arl8b and JIP4-Dynein complexes to regulate lysosome size and positioning. *Nature Communications*, 13(1). <https://doi.org/10.21203/rs.3.rs-345822/v1>
28. Gaurav Kumar, **Prateek Chawla, Neha Dhiman**, Sanya Chadha, Sheetal Sharma, Kanupriya Sethi, **Mahak Sharma**, and Amit Tuli (2022). RUFY3 links Arl8b and JIP4-Dynein complex to regulate lysosome size and positioning. *Nature Communications*, 13(1), 1540 .<https://doi.org/10.1038/s41467-022-29077-y>
29. Gokul G and **Jogender Singh (2022)**. Dithiothreitol causes toxicity in *C. elegans* by modulating the methionine-homocysteine cycle. *eLife*, 11(1), 76021. <https://doi.org/10.7554/eLife.76021>
30. Janice D. Pata, Y. Whitney Yin and **Indrajit Lahiri (2022)**. Editorial: Nucleic Acid Polymerases: The Two-Metal-Ion Mechanism and Beyond. *Frontiers in Molecular Biosciences*, 9(1), 948326. <https://doi.org/10.3389/fmolsb.2022.948326>
31. Jasreen Kaur, **Sharvan Sehrawat**, Ikjot Singh Sohal, Harpreet Singh, Naveen Kumar Gupta, Sanjeev Puri, Dhimiter Bello, and D. Madhu Khatri (2022). Toxicity screening and ranking of diverse engineered nanomaterials using established hierarchical testing approaches with a complementary in vivo zebrafish model. *Environmental Science: Nano*, 9(8), 2726-2749. <https://doi.org/10.1039/d2en00265e>
32. Jatin Chadha, Ravi, **Jogender Singh**, Sanjay Chhibber, and Kusum Harjai(2022). Gentamicin Augments the Quorum Quenching Potential of Cinnamaldehyde In Vitro and Protects *Caenorhabditis elegans* From *Pseudomonas aeruginosa* Infection. *Frontiers in Cellular and Infection Microbiology*, 12(1), 899566. <https://doi.org/10.3389/fcimb.2022.899566>

33. **Jayati Gera, Prerna Budakoti, Meghna Suhag, Lolitika Mandal and Sudip Mandal (2022).** Physiological ROS controls Upd3-dependent modeling of ECM to support cardiac function in Drosophila. *Science Advances*, 8(7). <https://doi.org/10.1126/sciadv.abj4991>
34. **Jayati Gera, Prerna Budakoti, Meghna Suhag, Lolitika Mandal, and Sudip Mandal (2022).** Physiological ROS controls Upd3-dependent modeling of ECM to support cardiac function in Drosophila. *Science Advances*, 8(7), 1-15. <https://doi.org/10.1126/sciadv.abj4991>
35. **Jillella Mallikarjun, and J. Gowrishankar(2022).** Essential Role for an Isoform of Escherichia coli Translation Initiation Factor IF2 in Repair of Two-Ended DNA Double-Strand Breaks. *Journal of Bacteriology*, 204(4), 00571-21. <https://doi.org/10.1128/jb.00571-21>
36. **Jillella Mallikarjun, L. SaiSree, P. Himabindu, K. Anupama, Manjula Reddy, and J. Gowrishankar (2022).** Modulation of RecFORQ- and RecA-Mediated Homologous Recombination in Escherichia coli by Isoforms of Translation Initiation Factor IF2. *Journal of Bacteriology*, 204(4), 569-21. <http://doi.org/10.1128/jb.00569-21>
37. John P. Gillies, Janice M. Reimer, Eva P. Karasmanis, **Indrajit Lahiri**, Zaw Min Htet, Andres E. Leschziner and Samara L. Reck-Peterson (2022). Structural basis for cytoplasmic dynein-1 regulation by Lis1. *Elife*, 11(6), 71229. <https://doi.org/10.7554/elife.71229>
38. **Kajal Gupta, Gaganpreet Kaur, Tejal Pathak, and Indranil Banerjee (2022).** Systematic review and meta-analysis of human genetic variants contributing to COVID-19 susceptibility and severity. *Gene*, 844(1), 146790. <https://doi.org/10.1016/j.gene.2022.146790>
39. **Karan Singh, Manas Arun Samant and Nagaraj Guru Prasad (2022).** Evolution of cross-tolerance in Drosophila melanogaster as a result of increased resistance to cold stress. *Scientific Reports*, 12(1), 19536. <https://doi.org/10.1038/s41598-022-23674-z>
40. **KomalaMaggu, SnehaKapse, Neetika Ahlawat, Manasa Geeta Arun, and Nagaraj Guru Prasad (2022).** Finding love: fruit fly males evolving under higher sexual

selection are inherently better at finding receptive females. *Animal Behaviour*, 187(1), 15–33. <https://doi.org/10.1016/j.anbehav.2022.02.010>

41. Kshitish C. Majumdar and **Rajesh Ramachandran**(2022). Aquaculture Productivity Enhancement Through Advanced Technologies. *Advances in Fisheries Biotechnology*, 1-28. [https://doi.org/10.1007/978-981-16-3215-0\\_1](https://doi.org/10.1007/978-981-16-3215-0_1)
42. **Kusum Lata, Mahendra Singh, Shamaita Chatterjee and Kausik Chattopadhyay**(2022). Membrane Dynamics and Remodelling in Response to the Action of the Membrane-Damaging Pore-Forming Toxins. *The Journal of Membrane Biology*, 255(2-3), 161–173. <https://doi.org/10.1007/s00232-022-00227-z>
43. **Lisha Arora, and Samrat Mukhopadhyay (2022)**. Conformational Characteristics and Phase Behavior of Intrinsically Disordered Proteins—Where Physical Chemistry Meets Biology. *Journal of Physical Chemistry B*, 126(28), 5137-5139. <https://doi.org/10.1021/acs.jpcb.2c04017>
44. **Mahendra Singh, N. Rupesh, Shashi Bhushan Pandit, andKausik Chattopadhyay (2022)**. Curcumin Inhibits Membrane-Damaging Pore-Forming Function of the  $\beta$ -Barrel Pore-Forming Toxin Vibrio cholerae Cytolysin. *Frontiers in Microbiology*, 12(1), 809782. <https://doi.org/10.3389/fmicb.2021.809782>
45. **Manas Geeta Arun, Tejinder Singh Chechi, Rakesh Meena, Shradha Dattaraya Bhosle, Srishti, and Nagaraj Guru Prasad (2022)**. Investigating the interaction between inter-locus and intra-locus sexual conflict using hemiclonal analysis in *Drosophila melanogaster*. *BMC Ecology and Evolution*, 22(1), 221992. <https://doi.org/10.1186/s12862-022-01992-0>
46. Matthieu Pierre Platret, **Santosh B. Satbhai**, Lukas Brent, Matias F. Gleason, Min Cao, Magali Grison, Marie Glavier, Ling Zhang, Christophe Gaillochet, Christian Goeschl, Marco Giovannetti, Balaji Enugutti, Julie Neveu, Marcel von Reth, Ruben Alcázar, Jane E. Parker, Grégory Vert, Emmanuelle Bayer, and Wolfgang Busch (2022). The receptor kinase SRF3 coordinates iron-level and flagellin dependent defense and growth responses in plants. *Nature Communications*, 13(1), 4445. <https://doi.org/10.1038/s41467-022-32167-6>

47. Mohammad Urfan, Haroon Rashid Hakla, Shubham Sharma, Manu Khajuria, **Santosh B. Satbhai**, Dhiraj Vyas, Sunil Bhougal, Narendra Singh Yadav, and Sikander Pal (2022). Paclobutrazol improves surface water use efficiency by regulating allometric trait behavior in maize. *Chemosphere*, 307(1), 2-13. <https://doi.org/10.1016/j.chemosphere.2022.135958>
48. **Neetika Ahlawat, Komal Maggu, Jigisha, Manas Geeta Arun, Abhishek Meena, Amisha Agarwala, and Nagaraj Guru Prasad (2022)**. No major cost of evolved survivorship in *Drosophila melanogaster* populations coevolving with *Pseudomonas entomophila*. *Proceedings of the Royal Society B: Biological Sciences*, 289(1974), 2-11. <https://doi.org/10.1098/rspb.2022.0532>
49. **Neetika Ahlawat, Manas Geeta Arun, Komal Maggu, Jigisha, Aparajita Singh, and Nagaraj Guru Prasad (2022)**. *Drosophila melanogaster* hosts coevolving with *Pseudomonas entomophila* pathogen show sex-specific patterns of local adaptation. *BMC Ecology and Evolution*, 22(1), 77. <https://doi.org/10.1186/s12862-022-02031-8>
50. **Nidhi Kumari**, and Lal Chand Rai (2022). Molecular characterization of local cyanobacterial isolates using 16S rRNA rpoB and nif H biomarkers. *Cyanobacterial Lifestyle and its Applications in Biotechnology*, 307-334. <https://doi.org/10.1016/B978-0-323-90634-0.00004-4>
51. **Paresh Nath Das, Aabeer Kumar Basuand Nagaraj Guru Prasad (2022)**. Increasing adult density compromises survival following bacterial infections in *Drosophila melanogaster*. *Journal of Insect Physiology*, 141(1), 104415. <https://doi.org/10.1016/j.jinsphys.2022.104415>
52. **Parvathy Ramesh, Sushmit Ghosh, and Lolitika Mandal(2022)**. Combination of Immunofluorescence and Quantitative Fluorescence In-situ Hybridization for Analysing Differential Gene Expression in the Niche Cells of the *Drosophila* Lymph Gland. *Bio-Protocol*, 12(2), 4290. <https://doi.org/10.21769/bioprotoc.4290>
53. **Prachi Ojha, Subhajit Pal and Samarjit Bhattacharyya (2022)**. Regulation of Metabotropic Glutamate Receptor Internalization and Synaptic AMPA Receptor Endocytosis by the Postsynaptic Protein Norbin. *The Journal of neuroscience*, 42(5), 731–748. <https://doi.org/10.1523/JNEUROSCI.1037-21.2021>

54. **Pratima Pandey**, Gazaldeep Kaur and **Kavita Babu** (2022). Crosstalk between neurons and glia through G-protein coupled receptors: Insights from *Caenorhabditis elegans*. *Progress in Molecular Biology and Translational Science*, 193(1), 119-144. <https://doi.org/10.1016/bs.pmbts.2022.06.005>
55. **Priyanka Dogra, Shruti Arya, Avinash K Singh, Anindya Datta, and Samrat Mukhopadhyay** (2022). Conformational and Solvation Dynamics of an Amyloidogenic Intrinsically Disordered Domain of a Melanosomal Protein. *The Journal of Physical Chemistry B*, 126(2), 443–452. <https://doi.org/10.1021/acs.jpcb.1c09304>
56. **R. Dutta, T. S. Chechi, A. Yadav, and N. G. Prasad** (2022). Indirect selection on cuticular hydrocarbon divergence in *Drosophila melanogaster* populations evolving under different operational sex ratios. *Journal of Zoology*, 316(3), 188-196. <https://doi.org/10.1111/jzo.12943>
57. Ravi Kumar Sharma, Jyoti Sharma, **Rajendra Kumar**, Darshan Badal, Ajinkya Pattekar, Shobha Sehgal, Amod Gupta, Pooja Jain, and Naresh Sachdeva(2022). TLR9 signalling activation via direct ligation and its functional consequences in CD4 + T cells. *Scandinavian Journal of Immunology*, 96(5), 2-18. <https://doi.org/10.1111/sji.13214>
58. Rohit Goswami and **Ruhila S.** (2022). High Throughput Reproducible Literate Phylogenetic Analysis. *PDGC 2022 - 2022 7th International Conference on Parallel Distributed and Grid Computing*, 337-340. <https://doi.org/10.1109/PDGC56933.2022.10053210>
59. Rohit Goswami, **Ruhila S.**, Amrita Goswami, Sonaly Goswami, and Debabrata Goswami (2022). Reproducible High Performance Computing without Redundancy with Nix. *PDGC 2022 - 2022 7th International Conference on Parallel Distributed and Grid Computing*, 238-242. <https://doi.org/10.1109/PDGC56933.2022.10053342>
60. **Rohit Kapila, Soumyadip Poddar, Neeraj Meena, and Nagaraj Guru Prasad** (2022). Investment in adult reproductive tissues is affected by larval growth conditions but not by evolution under poor larval growth conditions in *Drosophila*

melanogaster. ***Current Research in Insect Science***, 2(1), 100027.  
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61. Ruchira Sen, **Kunika Malhotra, Manisha Gupta, Rajbir Kaur, Divya Bawa, Meghna Duhan, Sonia Sandhi, Pratibha Songara, Deepak Nain, and Rhitoban Raychoudhury (2022)**. Coping with the ‘Indian summer’: unique nesting cycle and nest architecture of the paper wasp Polistes wattii. ***Science of Nature***, 109(3), 31. <https://doi.org/10.1007/s00114-022-01801-0>
62. Rupam Paul, Sourav Banerjee, Samarpita Sen, **Pratiksha Dubey, Saptarshi Maji, Anand K.Bachhawat, Rupak Datta, and Arnab Gupta (2022)**. A novel leishmanial copper P-type ATPase plays a vital role in parasite infection and intracellular survival. ***Journal of Biological Chemistry***, 298(2), 101539.  
<https://doi.org/10.1016/j.jbc.2021.101539>
63. **Samrat Mukhopadhyay (2022)**. Shapeshifting proteins: The role of structural disorder and conformational plasticity in physiology and disease. ***Essays in Biochemistry***, 66(7), 817-819. <https://doi.org/10.1042/EBC20220197>
64. **Samrat Mukhopadhyay, Aishwarya Agarwal, Sandeep K.Rai, Anamika, Avni, and Lisha Arora(2022)**. Aberrant phase transitions of a pathological Stop codon mutant of the prion protein. ***Biophysical Journal***, 121(3), 473a.  
<https://doi.org/10.1016/j.bpj.2021.11.420>
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<https://doi.org/10.1016/j.bios.2022.113995>
67. Savita Budania, **Abhishek Dubey, and Ajit Singh (2022)**. Trypanosoma evansiRoTat 1.2 variant surface antigen mimotopes selected by panning of the random peptide

phage-display library against monoclonal antibodies. *Journal of Molecular Recognition*, 35(11), 2984. <https://doi.org/10.1002/jmr.2984>

68. **Sayanta Mahapatra, Anusha Sarbahi**, Priyanka Madhu, Hema M. Swasth, Abhishek Sharma, Priyanka Singh, and **Samrat Mukhopadhyay** (2022). Substoichiometric Hsp104 regulates the genesis and persistence of self-replicable amyloid seeds of Sup35 prion domain. *Journal of Biological Chemistry*, 298(8), 102143. <https://doi.org/10.1016/j.jbc.2022.102143>
69. **Sayantan Goswami** and **Jayaraman Gowrishankar** (2022). Role for DNA double strand end-resection activity of RecBCD in control of aberrant chromosomal replication initiation in Escherichia coli. *Nucleic Acids Research*, 50(15), 8643-8657. <https://doi.org/10.1093/nar/gkac670>
70. **Shalini Rawat, Dhruba Chatterjee, Rituraj Marwaha, Gitanjali Charak**, Gaurav Kumar, **Shrestha Shaw, Divya Khatter**, Sheetal Sharma, Cecilia de Heus, Nalan Liv, Judith Klumperman, Amit Tuli, and **Mahak Sharma** (2022). RUFY1 binds Arl8b and mediates endosome-to-TGN CI-M6PR retrieval for cargo sorting to lysosomes. *Journal of cell biology*, 222(1), 2108001. <https://doi.org/10.1083/jcb.202108001>
71. **Shashi Prakash Yadav, Sanjeev Routh**, and **ArunikaMukhopadhyaya** (2022). Toll-Like Receptors (TLRs): Host immunity and bacterial ligand recognition. *Advances in Health and Disease*. 58(1), 1-47. <https://novapublishers.com/shop/advances-in-health-and-disease-volume-58/>
72. Sonika Chibh, **Shradha Suyal**, Nidhi Aggarwal, **Anand Kumar Bachhawat** and Jiban Jyoti Panda (2022). Cysteine-phenylalanine-derived self-assembled nanoparticles as glutathione-responsive drug-delivery systems in yeast. *Journal of Materials Chemistry B*, 10(42), 8733-8743. <https://doi.org/10.1039/d2tb01362b>
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74. **Sudhakar Singh, Surbhi Dahiya, Yuviana J Singh, Komal Beeton, Ayush Jain, Roman Sarkar, Abhishek Dubey, Azeez Tehseen, and SharvanSehrawat (2022).** Robust anti-SARS-CoV2 single domain antibodies cross neutralize multiple viruses. *iScience*, 25(7), 104549. <https://doi.org/10.1016/j.isci.2022.104549>
75. Suman Mishra, Ishika Pramanick, Anil Kumar, Somnath Dutta, **Nidhi Kundu, and Kausik Chattopadhyay (2022).** Structural insights into thermostable direct hemolysin of *Vibrio parahaemolyticus* using single-particle cryo-EM. *Proteins-Structure Function and Bioinformatics*, 91(2), 137-146. <https://doi.org/10.1002/prot.26416>
76. **Surbhi Dahiya, Sudhakar Singh, and SharvanSehrawat (2022).** Protocol for investigating the biogenesis of SARS-CoV-2 S pseudoviruses in HEK293T cells transduced to express the virus-specific intrabodies. *STAR Protocols*, 4(1), 1-14. <https://doi.org/10.1016/j.xpro.2022.101977>
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